Open Access

Clinical value of 3'-deoxy-3'-[¹⁸F] fluorothymidine-positron emission tomography for diagnosis, staging and assessing therapy response in lung cancer

Bandar Alwadani^{1,2}, Sergio Dall'Angelo^{2†} and Ian N. Fleming^{2*†}

Abstract

Lung cancer has the highest mortality rate of any tumour type. The main driver of lung tumour growth and development is uncontrolled cellular proliferation. Poor patient outcomes are partly the result of the limited range of effective anti-cancer therapies available and partly due to the limited accuracy of biomarkers to report on cell proliferation rates in patients. Accordingly, accurate methods of diagnosing, staging and assessing response to therapy are crucial to improve patient outcomes. One effective way of assessing cell proliferation is to employ non-invasive evaluation using 3'-deoxy-3'-[¹⁸F]fluorothymidine ([¹⁸F]FLT) positron emission tomography [¹⁸F]FLT-PET. [¹⁸F]FLT, unlike the most commonly used PET tracer [¹⁸F]fluorodeoxyglucose ([¹⁸F]FDG), can specifically report on cell proliferation and does not accumulate in inflammatory cells. Therefore, this radiotracer could exhibit higher specificity in diagnosis and staging, along with more accurate monitoring of therapy response at early stages in the treatment cycle. This review summarises and evaluates published studies on the clinical use of [¹⁸F]FLT to diagnose, stage and assess response to therapy in lung cancer.

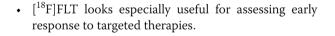
Keywords: Positron emission tomography, PET, Lung cancer, Fluorothymidine, [¹⁸F]FLT

Key points

- [¹⁸F]FLT is a useful surrogate biomarker of cell proliferation.
- [¹⁸F]FLT has higher specificity than [¹⁸F]FDG for diagnosing or staging lung cancer.
- [¹⁸F]FLT has lower sensitivity than [¹⁸F]FDG for diagnosing or staging lung cancer.
- [¹⁸F]FLT has good predictive values for assessing response to radiotherapy or chemoradiotherapy.

 $^{\rm 2}$ Institute of Medical Sciences, University of Aberdeen, Aberdeen AB25 2ZD, UK

Full list of author information is available at the end of the article



Background

Lung cancer (LC) remains the leading cause of cancerrelated mortality worldwide, accounting for about 18.4 % of all cancer cases in 2018. It ranks first in mortality and incidence in men. In women it has the second highest mortality and the third highest incidence [1]. LC is subcategorised histologically into small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC), of which adenocarcinoma (ADC), squamous cell carcinoma (SqCC) and large cell lung carcinoma (LCC) represents almost 80 % of diagnosed cases [2]. These subcategories show different growth patterns and might be associated with different prognoses [3].



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

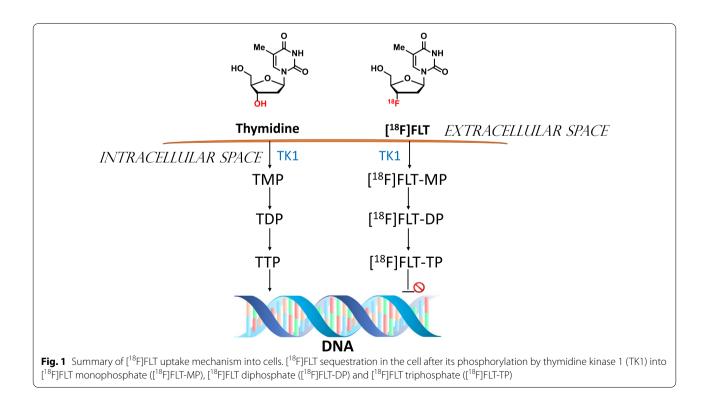
^{*}Correspondence: i.n.fleming@abdn.ac.uk

 $^{^{\}dagger}\text{Sergio}$ Dall'Angelo and Ian N. Fleming have contributed equally to this work

NSCLC can be stratified according to the tumour size, nodal involvement and metastases (TNM). SCLC is usually divided into limited or extensive disease, although the TNM system has been adopted by some clinical committees. These models, such as those provided by the Veterans Administration Lung Study Group and later modified by the International Association for the Study of Lung Cancer (IASLC), have proven useful to guide prognosis and patient stratification for therapy [4].

Uncontrolled cell proliferation is a fundamental hallmark of malignant tumour growth. The biomarker Ki-67 is considered the gold standard for assessment of cell proliferation due to the strong correlation between the cell proliferation rate and Ki67 expression in cells [5, 6]. However, it exhibits several potential drawbacks in the clinic, including the requirement for invasive collection of biopsies and possible sampling bias, due to poorly representative biopsies collected from heterogeneous tumours [7]. Over the past few decades, noninvasive positron emission tomography (PET) imaging has played an increasingly important role in LC management. [¹⁸F]Fluorodeoxyglucose ([¹⁸F]FDG) is the most widely used radiotracer in PET. It is a glucose analogue which monitors glucose metabolism, based on the concept that tumour cells take up significantly higher levels of glucose than normal tissues. It has been a beneficial adjunct in characterisation of intermediate solitary lung nodules and pre-treatment detection and staging of distant metastases [8]. However, [¹⁸F]FDG uptake is modulated by multiple signalling pathways, so is not selective enough to specifically assess changes in the cell proliferation rate. Moreover, a meta-analysis of [¹⁸F]FDG in LC showed its extreme heterogeneity in LC in pulmonary areas with inflammations [9].

The first PET radiotracer introduced for in vivo proliferative imaging was ¹¹C-labelled thymidine ([¹¹C]thymidine); however, its rapid degradation with a half-life time of only 20 min is a major limitation [10]. 3'-deoxy-3'-[¹⁸F]fluorothymidine ([¹⁸F]FLT) was therefore subsequently employed, as this ¹⁸F-labelled thymidine analogue has a half-life of about 110 min. [¹⁸F]FLT is initially phosphorylated by cytosolic thymidine kinase 1 (TK1) into FLT-monophosphate and then subsequently further phosphorylated to make the diphosphate and triphosphate nucleotide (Fig. 1). TK1 is the principal enzyme that controls the rate of nucleotide recycling via the salvage pathway of DNA synthesis. After [¹⁸F] FLT is phosphorylated to the triphosphate analogue it cannot be further metabolised or incorporated into the DNA molecule. It is therefore trapped intracellularly as its high hydrophilicity means that it cannot readily cross the cell membrane. This cell sequestration of [¹⁸F]FLT is due to the substitution of the hydroxyl group at the 5'-end of thymidine, which is essential for



ligation of DNA, with a fluorine-18 radionuclide. TK1 expression levels increase dramatically in proliferating cells, and there is close correlation between expression of the enzyme and the cell proliferation rate [11, 12]. Close correlation has also been observed between Ki-67 scoring and [¹⁸F]FLT uptake in many tumour types [5], including LC [6]. Therefore, $[^{18}F]FLT$ is a very plausible alternative to Ki-67 for assessing the proliferation rate in tumours. It also offers the advantages of avoiding the need to collect invasive biopsies and allows evaluation of proliferation heterogeneity across the entire tumour, minimising both patient discomfort and sampling errors. Moreover, [¹⁸F]FLT-PET is able to produce repeated 3D images for multiple cancer sites simultaneously, which is a distinct advantage for accurately assessing response to therapy. Alternative proliferation radiotracers have been also developed as direct biomarkers for DNA synthesis as they can be incorporated into DNA. This includes 2'-deoxy-2'-fluoro- $([^{18}F]FMAU)$ 5-methyl-1-β-D-arabinofuranosyluracil 1-(2-deoxy-2-[¹⁸F]fluoro-β-D-arabinofuranosyl)and 5-bromouracil ([¹⁸F]FBAU). However, these radiotracers are poor substrates for TK1 and show low uptake in highly proliferating tissue compared with [¹⁸F]FLT. This probably reflects their phosphorylation by mitochondrial thymidine kinase 2 (TK2) rather than TK1 [13], making [¹⁸F]FLT potentially superior to other proliferation radiotracers.

Relatively similar imaging protocols are followed in most [¹⁸F]FLT-PET scans. No specific preparations are required by patients although it is advantageous if they are instructed to drink up to 1 L water before imaging to stimulate tracer excretion from the renal calyces [14, 15]. The tracer dose administered ranges from 130 to 550 MBq. Most protocols employ static scans, although some more involved studies use dynamic scans to obtain more complex datasets. Subtly different scan timings and imaging parameters such as field of view and reconstruction techniques are also performed. For PET/CT, the patient is usually positioned supine, with arms raised to minimise beam hardening. Image evaluation is usually performed by two or more nuclear medicine physicians blinded to patient's data.

The aim of this literature review paper is to review the clinical value of [¹⁸F]FLT-PET proliferative imaging for diagnosis, staging and assessing response to therapy in LC. A secondary objective is to analyse the uptake values of [¹⁸F]FLT-PET in the various main LC sub-types in comparison with [¹⁸F]FDG and whether there is any difference between the two tracers in these sub-types. This overview will further enable clinicians to better understand the added value of [¹⁸F]FLT for clinical

management of LC and its proliferative pattern in comparison with [¹⁸F]FDG in different LC sub-types.

Materials and methods

Search strategy

To identify all relevant publications, a systematic search of Scopus/PubMed databases was implemented from inception to 1st December 2020 using combinations of the following keywords: "positron emission tomography", "PET", "lung cancer", "fluorothymidine" and "[¹⁸F]FLT".

Selection process

All potentially relevant publications were screened for eligibility. Initially, titles and abstracts were screened and if necessary, the full texts were scrutinised as well. Studies were included if they met the following criteria:

- It was an original study that investigated the performance of [¹⁸F]FLT-PET for diagnosing, staging or assessing therapy response in LC patients;
- 2. It involved patients with suspected or confirmed malignant lesions;
- 3. The patients underwent chemotherapy, radiotherapy, chemoradiotherapy or targeted therapy; and
- 4. Clinical outcome was assessed.

Studies were excluded if they:

- 1. Only involved animal or in vitro studies;
- 2. Were not written in English or were not accessible in full text; and
- 3. Involved certain publication types: case reports, reviews, legal cases, editorials, letters, interviews, and comments.

Statistical analysis of [¹⁸F]FLT uptake versus [¹⁸F]FDG uptake in lung cancer sub-types

Statistical analysis was performed using SPSS software. The specific research questions posed were as follows:

Is there any difference between the mean [¹⁸F]FDG and [¹⁸F]FLT uptake values in lung cancer patients? Null hypothesis was that there is no difference. Analysis was done using an unpaired students t-test.

Is there any different in mean radiotracer uptake values between the different lung cancer sub-types? This test was performed independently for [¹⁸F]FDG and [¹⁸F]FLT. Null hypothesis was that there is no difference. Statistical analysis was performed using a 1-way ANOVA with tukeys post hoc analysis and bootstrapping.

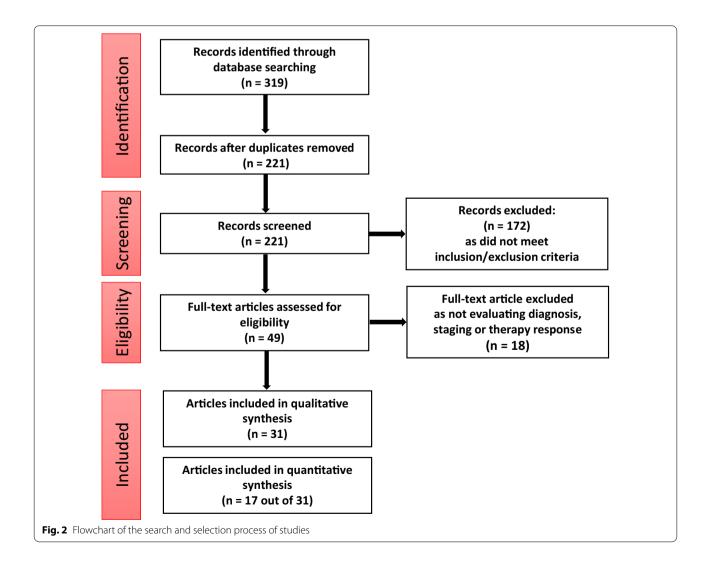
Results

Search results

The literature search identified a total of 319 studies: 210 in Scopus and 109 in PubMed. After removing the studies that were found in both databases, 221 studies remained. Figure 2 presents the flow chart of the search and selection procedure. Out of 221, only 31 studies were included as eligible, and out of these 31 studies, 17 studies were included for quantitative synthesis. A summary of [¹⁸F]FLT-PET LC clinical studies and their findings is listed in Table 1 and 2.

Diagnosis studies

The past two decades have witnessed a growing attention to $[^{18}F]FLT$ as a potential diagnostic to aid LC patient management. Imaging with $[^{18}F]FLT$ has shown added benefit to diagnostic accuracy. Buck et al. (n = 30) and Vessel et al. (n = 10) introduced the clinical potential of $[^{18}F]FLT$ in LC by demonstrating that the tracer accumulates primarily in malignant lesions, with minimal uptake in benign lesions. The latter study also showed ^{[18}F]FLT maximum standardised uptake value (SUVmax) strongly correlates with both Ki-67 index and S-phase fraction (SPF), measured using flow cytometry [14, 16]. Further, Buck et al. observed mean SUVmax of 5.2 in malignant tumours compared with no apparent uptake in benign tumours [16]. This observation was supported by another study which showed primary tumours exhibiting almost identical [¹⁸F]FLT uptake values [17]. Furthermore, Wang et al. (n = 55) observed significantly higher $[^{18}F]$ FLT uptake (p < 0.05) in LC lesions compared with all other solitary pulmonary nodules, including tuberculosis, inflammatory and benign lesions, whereas tuberculosis showed similar [¹⁸F]FDG-SUVmax (6.9) values to LC (6.8) [18]. In multiple comparative studies with $[^{18}F]$ FDG [18-21], higher specificity (ability to exclude nonmalignant lesions) was presented using [18F]FLT. The imaging specificity of [¹⁸F]FLT ranges from about 77% to



99% versus about 50% to 84% for imaging with [¹⁸F]FDG in a total number of 195 patients of all study cohorts combined.

On the other hand, the relatively low uptake of $[^{18}F]FLT$ adversely affects its detection ability in comparison with ^{[18}F]FDG. For example, Buck et al. exhibited the significantly lower uptake of [18F]FLT compared with [18F]FDG (p < 0.05) [17]. Similar significant differences between [¹⁸F]FLT and [¹⁸F]FDG uptakes were also reported in a number of other studies [18-20, 22-26]. Also, lower sensitivity was shown with [18F]FLT-PET compared with [¹⁸F]FDG-PET. The sensitivity of [¹⁸F]FLT-PET range was only 65%-83% versus 85%-97% for [¹⁸F]FDG-PET in 195 patients combined from the mentioned studies. Given the higher specificity of [18F]FLT-PET but lower sensitivity in comparison with [¹⁸F]FDG-PET, it was proposed that a combination of both [18F]FLT and [18F]FDG-PET could potentially provide better diagnostic performance than the individual tracers. Indeed, this strategy was conducted by a multicentre study consisted 55 patients with suspected LC [21], resulting in sensitivity improving from 87% (with [18F]FDG alone) to 100% and specificity increasing from 77% (with [¹⁸F]FLT alone) to approximately about 90%.

Proliferative imaging of lung cancer sub-types with [¹⁸F] FLT versus [¹⁸F]FDG

Histological uptake in this review showed that [¹⁸F] FLT-SUVmax is significantly lower than [¹⁸F]FDG-SUVmax across all LC histotypes (p < 0.0001), with different uptake values in the various histotypes. [¹⁸F]FDG-SUVmax, as illustrated in Fig. 3, demonstrated the greatest mean SUVmax in SCLC (12.03) followed by SqCC (9.38), LCC (8.60), and ADC (8.10). A comparison of [¹⁸F]FDG-SUVmax detected no significant difference in the 4 subtypes of LC (p > 0.05), although the statistical power of this analysis is limited by the number of SCLC studies performed to date.

The highest [¹⁸F]FLT-mean SUVmax was in SqCC (4.65) followed by LCC (4.07), ADC (3.82), and SCLC (2.39). A comparison of [¹⁸F]FLT-SUVmax between the different histotypes observed that [¹⁸F]FLT-SUVmax was significantly lower in SCLC than in SqCC (p < 0.05). Uptake patterns with both tracers in NSCLC sub-types are consistent with the proliferative patterns demonstrated by Ki-67 in a previous study in the different sub-types of NSCLC histology [26].

Staging studies

Staging of LC is another clinical application that has been evaluated in several studies. In general, [¹⁸F]FLT-PET has failed to demonstrate better TNM staging than [¹⁸F] FDG-PET. Initially, a preliminary study in 2004 showed

that, in comparison with clinical TNM staging, [¹⁸F] FLT may be limited; [¹⁸F]FLT staging identified 9 out of 17 patients incorrectly [15]. Buck et al. compared $[^{18}F]$ FLT-PET with [¹⁸F]FDG-PET for staging 47 patients with suspected malignant nodules. The clinical TNM stage according to histopathology was correctly identified in 67% of [18F]FLT scans compared with 85% of [18F]FDG scans [17]. Furthermore, Yap et al. (n = 22) also evaluated ^{[18}F]FLT staging in comparison with ^{[18}F]FDG, taking histopathology data as a reference standard. According to histopathology, 3 patients were disease free, 10 patients were at early resectable stages and 7 were at late inoperable stages. [¹⁸F]FLT overstaged two patients (9%) and understaged eight patients (36%) whereas [¹⁸F]FDG overstaged six patients (27%) and understaged three patients (14%) [46]. Likewise, Yang et al. (n = 31) observed that more patients are understaged with [¹⁸F]FLT and more patients are overstaged with [18F]FDG. In this study, understaged patients represent 16% with [18F]FLT and 6% with [¹⁸F]FDG and overstaged patients represent 6% with [¹⁸F]FLT and 16% with [¹⁸F]FDG [20].

In addition, sensitivity, specificity and accuracy of [¹⁸F] FLT for nodal involvement were evaluated in 5 studies in comparison with [¹⁸F]FDG. The data showed clear differences between the two tracers. Buck et al. (n = 47) compared [18F]FLT-PET staging with [18F]FDG-PET staging in 47 patients with suspected malignant nodules and found that the clinical TNM stage was correctly identified in 67% of [¹⁸F]FLT scans compared with 85% of [¹⁸F] FDG scans [17]. Although both [¹⁸F]FLT and [¹⁸F]FDG showed 100% specificity for staging of lymph nodes in this study, the sensitivity and accuracy of [18F]FDG was higher (77% and 83%) than those of [18F]FLT (53% and 67%). Another study of 31 NSCLC patients [20] demonstrated even greater specificity and accuracy for [¹⁸F]FLT in staging lymph nodes than [¹⁸F]FDG but lower sensitivity than that of [18F]FDG. The sensitivity, specificity and accuracy for [¹⁸F]FLT were 65%, 85% and 93%, respectively, whereas the equivalent $[^{18}F]FDG$ values were 98%, 84% and 84%, respectively [20]. In contrast, Yamamoto et al. (n = 34) showed the same sensitivity (57%) for both tracers but higher specificity and accuracy (93% and 85%) for [¹⁸F]FLT versus 78% and 74% for [¹⁸F]FDG [19]. Moreover, Xu et al. (n = 14) reported better sensitivity, specificity and accuracy for [¹⁸F]FLT (85%, 93% and 85%, respectively) than for [¹⁸F]FDG (93%, 78% and 84%) [26]. However, taken together, the weighted mean for sensitivity, specificity and accuracy in all of these studies for $[^{18}F]$ FLT are 61%, 94%, and 80%, respectively, and for $[^{18}F]$ FDG are 79%, 88% and 80%, respectively.

lable I summ	ary or clinica	il studies evaluating diagnosis (and staging and their iind	lings in lung car	able 1 summary of clinical studies evaluating diagnosis and staging and their inidings in lung cancer based on Larjerla-PET proliferative imaging	ive imaging	
Reference	۲	[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ F]FLT/[¹⁸ F]FDG different LC histotypes uptake (SUVmax)	Purpose	TNM stage	Reference standard	Findings
Buck et al. 2002 [16]	90 8	Mean SUVmax=5.2	SCLC (n = 1): 2.4	Diagnosis	$ \begin{array}{l} TxN0M0 \ (n=1); TxN0M1 \ (n=4); \\ T1N0M0 \ (n=2); T1N1M0 \ (n=2); \\ T2N0M0 \ (n=1); T2bN0M0 \ (n=1); \\ T2N1M0 \ (n=4); T2N2MX \ (n=1); \\ T2N2M1 \ (n=1); T3N0M0 \ (n=1); \\ T3N3M0 \ (n=1); T4N1M0 \ (n=1); \\ T4N2M0 \ (n=2); \end{array} $	Histopathology Ki-67	[¹⁸ F]FLT uptake was weak but easily detectable in Primary lesions. [¹⁸ F]FLT is strongly correlated with Ki-67 in the malig- nant lesions
Vesselle et al. 2002 [27]	10	SUVmean: 3.84	LCC $(n = 4)$: 5.29 SqCC $(n = 3)$: 3.58 ADC $(n = 2)$: 4.01	Diagnosis	NA	Histopathology Ki-67 S-phase fraction (SPF)	[¹⁸ F]FLT SUVmax was correlated with Ki-67 and SPF ($r = 0.78$, $p = 0.0043$) and ($r = 0.69$, $p = 0.03$), respectively
Dittmann et al. 2003 [23]	9	[¹⁸ F]FLT SUVmean: 4.0 [¹⁸ F]FDG SUVmean: 6.9	ΥZ	Diagnosis	Ą	Histopathology Ki-67 [¹⁸ FJFDG	[¹⁸ F]FDG uptake was significantly higher than [¹⁸ F]FLT uptake (ρ = 0.0006). [¹⁸ F]FLT accurately detected thoracic lesions. Detec- tion of lesions in the liver and the bone marrow was hampered with high physiological [¹⁸ F] FLT uptake
Buck et al. 2003 [22]	26	[¹⁸ FJELT SUVmean: 1.8 [¹⁸ FJFDG SUVmean: 4.1	[¹⁸ FJFLT: SCLC (N = 1): 2.4 [¹⁸ FJFDG: SCLC (N = 1): 12.7	Diagnosis	TxN0M0 (n = 1); TxN0M1 (n = 4); T1N0M0 (n = 1); T1N1M0 (n = 2); T2bN0M0 (n = 1); T2N1M0 (n = 4); T2N2Mx (n = 1); T3N0M0 (n = 1); T3N3M0 (n = 1); T4N2M0 (n = 2)	Histopathology Ki-67 [¹⁸ FJFDG	[¹⁸ FJFDG uptake was significantly higher than [¹⁸ FJFLT uptake (<i>p</i> =0.05)
Cobben et al. 2004 [15]	71	[¹⁸ F]FLT Median SUVmax: 2.7 [¹⁸ F]FDG Median SUVmax: 8	[¹⁸ FJFLT: ADC (n=4): 2.73 SqCC (n=4): 2.55 [¹⁸ FJFDG: ADC (n=4): 7.38 SqCC (n=4)=9.83	Diagnosis Staging	TxN2/3M1 (n = 1); T1N0M1 (n = 1); T2N0M0 (n = 1); T2N2M0 (n = 1); T2N2M1 (n = 1); T2N2/3M1 (n = 1); T2N3M1 (n = 1); T4N0M1 (n = 2); T4N1M1 (n = 1); T4N2M0 (n = 3); T4N2M1 (n = 3)	Histopathology [¹⁸ FJFDG	$[^{18}F]FDG$ uptake was significantly higher than $[^{18}F]FLT$ uptake (p = 0.012) Staging by $[^{18}F]FLT-PET was correctfor 8 of 17 patients$
Buck et al. 2005 [17]	47	[¹ 8F]FLT SUVmean: 3.7 [¹ 8F]FLT SUVmax: 5.5 [¹ 8F]FDG SUVmean: 6 [¹ 8F]FDG SUVmax: 11	[¹⁸ FJFLT: ADC (n=8): 4.34 SqCC (n=6): 6 LCC (n=1): 5.7 SCLC (n=1): 2.4 [¹⁸ FJFDG: ADC (n=8): 8.11 SqCC (n=6): 6.5 LCC (n=1): 12.5 SCLC (n=1): 12.7	Diagnosis Staging	TcisNOMO (n = 1); T1NOMO (n = 2); T1N1MO (n = 2); T1N3MO (n = 1); T2NOMO (n = 2); T2N1MO (n = 5); T2N2MO (n = 2); T3NOMO (n = 1); T3N3MO (n = 1); T4N1MO (n = 1); T4N2MO (n = 2); T4N3MO (n = 1)	Histopathology [¹⁸ FJFDG	$[^{18}F]FDG$ uptake was significantly higher than $[^{18}F]FLT$ uptake ($p = 0.05$). Clinical TNM stage was correctly identified in 67% of $[^{18}F]$ FLT scans, compared to 85% of $[^{18}F]FDG$ scans

Table 1 Summary of clinical studies evaluating diagnosis and staging and their findings in lung cancer based on [¹⁸FJFLT-PET proliferative imaging

	inea/						
Reference	۲	[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ FJFLT/[¹⁸ FJFDG different LC histotypes uptake (SUVmax)	Purpose	TNM stage	Reference standard	Findings
Yap et al. 2006 [24]	22	[¹⁸ FJFLT SUVmax: 1.6 [¹⁸ FJFDG SUVmax: 3.1	$\begin{bmatrix} {}^{18}FJFLT: ADC (n = 9): 1.11 \\ SqCC (n = 3): 1.7 \\ LCC (n = 1): 3.6 \\ \begin{bmatrix} {}^{18}FJFDG: ADC (n = 9): 1.87 \\ SqCC (n = 3): 3.53 \\ LCC (n = 1) = 7 \\ LCC (n = 1) = 7 \end{bmatrix}$	Diagnosis Stag- ing	0 (n=3); IA (n=5); IB (n=2); IIA (n=1); IIIA (n=2); IIIB (n=2); IV (n=7)	Histopathology Ki-67 [¹⁸ FJFDG	[¹⁸ FJFDG uptake was significantly higher than [¹⁸ FJFLT uptake (p < 0.05) [¹⁸ FJFLT overstaged (9%) of patients and under- staged (36%) of patients whereas [¹⁸ FJFDG over- staged (27%) of patients and understaged (14%) of patients
Yamamoto et al. 2007 [<mark>25</mark>]	18	(¹⁸ FJFLT SUVmean: 3.6 (¹⁸ FJFDG SUVmean: 8.5	$[^{18}F]FLT: ADC (n = 7): 3.43$ SqCC (n = 5): 6.26 $[^{18}F]FDG: ADC (n = 7): 7.46$ SqCC (n = 5) = 16.02	Diagnosis	T1N0M0 (n = 8); T1N2M0 (n = 1); T2N0M0 (n = 4); T2N1M0 (n = 2); T2N2M0 (n = 1); T3N2M0 (n = 1); T4N0M0 (n = 1)	Histopathology Ki-67 [¹⁸ FJFDG	$[^{18}F]FDG$ uptake was significantly higher than $[^{18}F]FLT$ uptake $(p < 0.0003)$
Tian et al. 2008 [21]	55	[¹⁸ F]FLT SUVmean: 3.54 [¹⁸ F] FDG SUVmean: 8.13	$[^{18}FJFLT: ADC (n = 10): 3.72SqCC (n = 2): 4.4[^{18}FJFDG: ADC (n = 10): 7.767.76SqCC (n = 2): 9$	Diagnosis with dual tracers [¹⁸ F]FDG/[¹⁸ F] FLT	₹Z	Histopathology [¹⁸ FJFDG RECIST	The sensitivity and specificity of [¹⁸ FJFDG and [¹⁸ FJFLT were 87.5% and 58.97% and 68.75% and 56.92%, respectively. Improved sensitivity and specificity, 100% and 89.74%, respectively were shown with dual tracer [¹⁸ FJFDG/[¹⁸ FJFLT
Yamamoto et al. 2008 [19]	6 4	(¹⁸ FJFLT SUVmean: 3.5 [¹⁸ FJFDG SUVmean: 11.4	[¹⁸ FJFLT: ADC (n = 16): 4.28 SqCC (n = 16): 5.57 SCLC (n = 2): 2.35 [¹⁸ FJFDG: ADC (n = 16): 9.88 SqCC (n = 16): 12.72 SCLC (n = 2): 10.7	Diagnosis Stag- ing	T1N1M0 (n = 1); T1N2M0 (n = 1); T2N0M0 (N = 10); T2N0M1 (N = 1); T2N1M1 (N = 2); T2N2M0 (N = 2); T3N0M0 (n = 1); T3N1M0 (n = 1); T3N2M0 (n = 2); T4N0M0 (n = 2);	Histopathology [¹⁸ FJFDG	$[^{18}$ FJFDG uptake was significantly higher than $[^{18}$ FJELT uptake ($P < 0.0001$). $[^{18}$ FJELT showed same sensitivity and higher specificity and accuracy than $[^{18}$ FJ FDG
Yang et al. 2010 [20]	E.	[¹⁸ FJFLT SUVmax: 4.2 [¹⁸ FJFDG SUVmax: 7.7	[¹⁸ FJFLT: ADC (n = 13): 3.8 SqCC (n = 11) = 5.7 [¹⁸ FJFDG: ADC (n = 13): 8.4 SqCC (n = 11) = 8.1	Diagnosis Stag- ing	T1N0 (n=4); T1N1 (n=5); T1N2 (n=2); T2N0 (n=6); T2N1 (n=4); T2N2 (n=2); T3N0 (n=3); T3N1 (n=3); T3N2 (n=2)	Histopathology [¹⁸ FJFDG	$[^{18}F]FLT$ showed significantly lower sensitivity ($p = 0.031$) for primary lesions. $[^{18}F]FLT$ showed better accuracy and specificity but lower sensitivity for nodal staging than $[^{18}F]FDG$

Table 1 (continued)	ued)						
Reference	۲	[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ FJFLT/[¹⁸ FJFDG different LC histotypes uptake (SUVmax)	Purpose	TNM stage	Reference standard	Findings
Xu et al. 2016 [26] 14	4	[¹⁸ F]FLT SUVmean: 4.96 [¹⁸ F] FDG SUVmean: 8.12	[¹⁸ FJFLT: ADC (n = 11): 5.14 Staging SqCC (n = 3): 4.28 [¹⁸ FJFDG: ADC (n = 11): 8.38 SqCC (n = 3): 7.13	Staging	T3N0M0 ($n = 4$); T3N1M0 ($n = 3$); T3N1M1 ($n = 2$); T4N0M0 ($n = 1$); T4N0M1 ($n = 1$); T4N1MX ($n = 1$); T4N1M0 ($n = 1$); T4N1M1 ($n = 1$)	Histopathology [¹⁸ FJFDG	[¹⁸ F]FDG uptake was significantly higher than [¹⁸ F]FLT uptake ($\rho = 0.01$). [¹⁸ F]FLT showed better accuracy and specificity but lower sensitivity for nodal stag- ing than [¹⁸ F]FDG
Wang et al. 2016 [18]	55	[¹⁸ FJFLT SUVmax: 2.9 [¹⁸ FJFDG SUVmax: 6.8	Ž	Diagnosis	Υ	Histopathology Ki-67 [¹⁸ FJFDG	[¹⁸ FJFDG uptake was significantly higher than [¹⁸ FJFLT uptake ($p < 0.001$) [¹⁸ FJFLT showed lower sensitivity (68.75%) compared with 87.8% for [¹⁸ FJEDG but higher secificity (77%) than [¹⁸ FJEDG (59%) [¹⁸ FJEDG (59%) tary pulmonary nodules [¹⁸ FJEDG showed an over- lap in detecting cancer and tuberculosis
All of the second to t	and the state	All of the consists is this table was sincely control of indice average at a					

All of the reports in this table were single centre studies except Tian et al. 2008 [21]

n; Number of patients, NA; Not available, LC; Lung cancer, SCLC; Small cell lung cancer, NSCLC; non-small cell lung cancer, ADC; Adenocarcinoma, SqCC; Squamous cell carcinoma, LCC; Large cell lung carcinoma, TNM; Tumour, nodal involvement and metastases, PET; Positron emission tomography, [¹⁸FJFLG; I¹⁸FJfluorodeoxyglucose, [¹⁸FJFLT; 3⁻deoxy-3⁻¹[¹⁸FJfluorothymidine, SUV; Standardised uptake value, SUVmean; Mean standardised uptake value, SUVmean; Mean standardised uptake value, SUVmean; Mean standardised uptake value, SUV; Standardised uptake value, SUVmean; Mean standardised uptake value, SUVmean; Mean

σ
gin
Jaç
.⊑.
≥
rati
life
oro
Ē
<u> </u>
Ë
Ē
[18
Ч
based
Cel
can
ğ
h
.⊆
gs
.≘
find
5
thei
đ
an
ISe
Ď
esp
Ž
rap
her
g tļ
tinč
uat
Val
S G
dies
stuc
_
ica
clin
of c
\geq
Ша
Ĩ
Su
2
Table
Tab
-

Reference	E	[¹⁸ F]FLT/[¹⁸ F]FDG LC primary Tumour uptake (SUV)	[¹⁸ FJFLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
Frings et al. 2013 [28]	14	Variable uptake	₹ Z	Study effect of pemetrexed- induced TS- inhibition on [¹⁸ F] FLT uptake 4 h after pemetrexed administration	ΨZ	Histopathology CT response	Changes in [¹⁸ F] FLT uptake 4 h after pemetrexed administration were not predictive for turmour response, TTP or OS
Crandall et al. 2017 [29]	6	Mean SUVmax: Base- line: 5.6 ± 2.0 Post-cycle 1 (day 15–21): 4.8 ± 2.5 Post-cycle 2 (day 36–42): 4.4 ± 2.3	۲ ۲	Study early chemotherapeu- tic response in comparison with [¹⁸ FJFDG	T1bN1M0 (n = 1); T1bN2M0 (n = 1); T2bN0M0 (n = 1); T2bN1M0 (n = 2); T3N0M0 (n = 2); T3N1M0 (n = 1); T3N2M0 (n = 1)	Histopathology CT response KI-67 [¹⁸ F]FDG	[¹⁸ FJFLT did not show significant difference between anatomic respond- ers and anatomic non-responders while [¹⁸ FJFDG showed significant difference between these groups
McHugh et al. 2018 [30]	4	SUVmax from baseline in two patients:—64.7% and—54.3%	A	Study the effect of dexamethasone on pemetrexed efficacy	АА	CT response	Decline of [¹⁸ FJFLT uptake from base- line, with a variable response between individual tumour lesions
Vera et al. 2011 [31]	Ŋ	Mean SUVmax: Baseline: 4.7 During therapy: 2.2	$[^{16}F]FLT: Baseline: ADC (n = 2): 4.64SqCC (n = 2): 3.86During therapy: ADC (n = 2): 3.86During therapy: ADC (n = 2): 2.13SqCC (n = 2): 2.13[^{18}F]FDG: Baseline: ADC (n = 2): 2.36SqCC (n = 2): 3.32ADC (n = 2): 3.32SqCC (n = 2): 5.38$	Study radiotherapy responses in com- parison with [¹⁸ F] FDG and F-miso	TXN3 (n = 1);T2N2 (n = 1);T3N2 (n = 1);T4N2 (n = 1);T4N3 (n = 1)	Histopathology	A significant decrease in SUVmax during radiotherapy was observed for $\Gamma^{18}F_{FDG}$ FLT and $\Gamma^{18}F_{FDG}$ but not for F-miso

Reference n	[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ F]FLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
Saga et al. 2011 [32] 20	Mean SUVmax: base- line: 1.54 3 months postther- apy: 0.35	₹ _Z	Longitudinal study of the responses to carbon-ion radiotherapy	T1N0M0 (n = 9); T2N0M0 (n = 11)	Histopathology CT response	[18 FJFLT uptake sig- niffcantly decreased after treatment Basline [18 FJFLT uptake of patients who developed recurrence and who died of LC were significantly higher than that of patients who did not (p = 0.008 and 0.007)
Trigonis et al. 2014 16 [33]	SUVmean: baseline: 2.2 ± 0.7 Response value: 1.6 ± 0.4 SUVmax: baseline: 5.3 ± 2.0 Response value: 4.1 ± 1.4	₹ Z	Study early radio- therapy response and test-retest variability	Ą	Histopathology CT response	SUVmean decreased by 25% in the absence of volumetric change (p = 0.0001) after 5-11 radiotherapy fractions. Larger decrease of 40% was shown in meta- static nodes with 31% decrease in volume (p < 0.0001). Similar findings for SUVmean reproducibility (standard deviation [SD]: 8.9%) in pri- mary tumours was better than SUVmax reproducibility (SD: 12.6%)

Reference n Everitt et al. 2009 5 [34]							
		[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ FJFLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
		SUVmax: mean reduc- tion of 0.58 × base- line	[¹⁸ FJFLT: Baseline: ADC (n = 1): 5.3 SqCC (n = 2): 8.25 LCC (n = 2): 5.15 [¹⁸ FJFDG: Baseline: ADC (n = 1): 15 SqCC (n = 2): 17.15 LCC (n = 2): 9.4	Establish [¹⁸ FJFLT assessment of cell proliferation during chemora- diotherapy	T2N0M0 (n = 1); T2N2M0 (N = 1); T2N3M0 (n = 1); T3N2M0 (n = 2)	Histopathology	Decline of [¹⁸ F] FLT uptake after chemotherapy in epithelial cancers and bone marrow (radiosensitive tissue)
		[¹⁸ FJFLT Median SUVmax: baseline: 6; wk2: 3; wk4: 2 [¹⁸ FJFDG Median SUVmax: baseline: 14; wk2: 10; wk4:10	۲Z	Study early chemo- radiotherapeutic response in comparison with [¹⁸ FJFDG	ΥN	Histopathology CT response	[¹⁸ FJELT is a more sen- sitive tracer for early treatment response than [¹⁸ FJFDG
Everitt et al. 2017 60 [36]	0	₹Z	¥ Z	Study relationship between chemo- radiotherapeutic responses and clinical outcomes in comparison with [¹⁸ FJFDG	₹	DG1 ^{[18}]	Stable uptake of [¹⁸ F] FLT at 2 wk after therapy was associ- ated with longer OS and PFS compared with patients whose tumours demon- strated reduced or absent [¹⁸ F]FLT uptake or absent [¹⁸ F]FLT uptake ciation between changes in [¹⁸ F] FLT uptake and clainical outcomes. This could be due to weakening tumourocidal effect of rediotherapy with inhibitory effect of possibly chemo- therapy (carboplatin and paclitaxel)

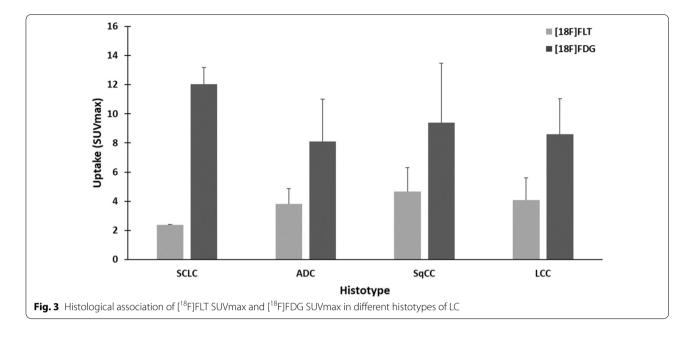
Table 2 (continued)						
Reference n	[¹⁸ FJFLT/[¹⁸ FJFDG LC primary Tumour uptake (SUV)	[¹⁸ FJFLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
Yang et al. 2012 [37] 68	Mean SUVmax: 4.1	Baseline: ADC (n = 30): 3.8 SqCC (n = 28): 5	Study antian- giogenic therapy responses in comparison with MVD	۶	Histopathology KI-67 MVD	[18 FJFLT uptake was significantly cor- related with MVD as reflected by CD105-MVD (r=0.633, $p = 0.000$). Patients with lower [18 FJ ELT uptake and CD105-MVD values had significantly higher median survival times than patients with higher [18 FJFLT uptake and CD105-MVD values ($p = 0.046$)
Scarpelli et al. 2018 14 [38]	Change in SUVmax: during angiogenic therapy: – 11% After combination chemother- apy: – 44%	٩	Study the sequen- tial responses of angiogenic therapy followed by chemotherapy	Ϋ́	Histopathology	[18 FJFLT uptake significantly decreased during therapy ($p = 0.04$) and after combina- tion chemotherapy ($p = 0.03$)
Scarpelli et al. 2018 33 [39]	A	ИА	Study the sequen- tial responses of angiogenic therapy followed by chemotherapy	٩	Histopathology	Tumour cell proliferation and vasculature were decreased 2 weeks after therapy and increased one week after therapy break
Sohn et al. 2008 [40] 31	Mean SUVmax decline, 7 days post- therapy:—36.0% for responders versus -10.1% for nonre- sponder (-10.9% was ued as cutoff)	Ą	Study early EGFR- TKI (gefitinib) therapy responses to predict clinical outcome	IV (n = 28)	CT response	[¹⁸ FJFLT uptake sig- nificantly decreased 7 days after gefitinib therapy in respond- ers. Responders had significantly longer TTP ($\rho = 0.03$) than non-responders

Table 2 (continued)							
Reference n		[¹⁸ F]FLT/[¹⁸ F]FDG LC primary Tumour uptake (SUV)	[¹⁸ F]FLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
Mileshkin et al. 2011 5' [41]	_	[¹⁸ FJFLT SUVmax: baseline: 2.5 [¹⁸ FJFDG SUVmax: 5.63	₹ Z	Study early EGFR- TKI (erlotinib) therapy responses to predict clini- cal outcome in comparison with [¹⁸ FJFDG	۲Z	CT response	PFS was predicted by both [¹⁸ F]FLT and [¹⁸ F]FDG 2wks and 8wks after therapy. OS was predicted by both tracers 8wks after therapy but by [¹⁸ F] FDG only 2wk after therapy
Zander et al. 2011 3 [42]	34	[¹⁸ FJFLT Mean SUVpeak: baseline: 3.47 week2: 3.12 ¹⁸ FJFDG Mean SUVpeak: baseline: 6.94 week2: 6.03	ę Z	Study early EGFR- TKI (erlotinib) therapy responses to predict clini- cal outcome in comparison with [¹⁸ FJFDG	Υ	CT response	[¹⁸ FJFLT predicted PFS but not OS 6 wks after therapy, while [¹⁸ FJFDG predicted PFS, OS 6 wks after therapy
Kahraman et al. 3 2011 [43]	30	ΨZ	۲	Study early EGFR- TKI (erlotinib) therapy responses to predict clini- cal outcome in comparison with [¹⁸ FJFDG	۲Z	CT response	Both [¹⁸ F]FLT and FDG predicted PFS 1wk and 6wk after therapy
Scheffler et al. 2013 4 [44]	0	Mean SUVmax,pretherapy: 3.0	$[^{18}$ FJFLT: Baseline: ADC (n = 29): 3.18 SqCC (n = 6): 3.53 LCC (n = 1): 1.9 LCC (n = 1): 1.9 [^{18}FJFDG: Baseline: ADC (n = 29): 7.09 SqCC (n = 6) = 6.38 LCC (n = 1): 6.6	Study the prog- nostic value of baseline uptake in patients treated with EGFR-TKI (erlotinib) in comparison with [¹⁸ FJFDG	۲Z	Histopathology Ki-67	[¹⁸ FJFLT to prognosti- cally stratify NSCLC patients treated with erlotinib as patients with low uptake had significanty longer survival times ($\rho = 0.027$) than patients with high uptake

Reference	£	[¹⁸ F]FLT/[¹⁸ F]FDG LC primary Tumour uptake (SUV)	[¹⁸ F]FLT/[¹⁸ F] FDG different LC histotypes uptake (SUVmax)	Purpose	TNM Stage	Reference standard	Findings
Bhoil et al. 2014 [45]	15	۲	M	Study TKI (gefitinib NA or erlotinib) therapy responses to predict clini- cal outcome in comparison with [¹⁸ FJFDG	M	CT response	Neither OS nor PFS were correlated with [¹⁸ FJFLT but both correlated with [¹⁸ FJFDG
All of the reports in n; Number of patier	this table involved sing its, NA; Not available, L	II of the reports in this table involved single centre studies except Crandall et al. 2017 [29] and Mileshkin et al. [41] ; Number of patients, NA; Not available, LC; Lung cancer, SCLC; Small cell lung cancer, NSCLC; non-small cell lung ca concernent in the more and independent of the development of the first of the state of the state of the state of	idall et al. 2017 [29] and N ell lung cancer, NSCLC; no Et T.3' Accounce, 2' (¹⁸ E-140.000)	Mileshkin et al. [41] on-small cell lung cancer othomiding SUV. Stands	All of the reports in this table involved single centre studies except Grandall et al. 2017 [29] and Mileshkin et al. [41] n; Number of patients, NA; Not available, LC; Lung cancer, SCLC; Small cell lung cancer, NSCLC; non-small cell lung cancer, ADC; Adenocarcinoma, SqCC; Squamous cell carcinoma, LCC; Large cell lung carcinoma, PET;	carcinoma, LCC; Large ce	ell lung carcinoma, PET;

Table 2 (continued)

which is a set of the set of



Studies assessing therapy response Chemotherapy

Three studies have investigated the potential of [¹⁸F]FLT for assessing chemotherapy and the findings were not promising. Frings et al. [28] showed that change in $[^{18}F]$ FLT uptake 4 hours after treatment with pemetrexed, an inhibitor of thymidylate synthase, was neither significantly correlated with time to progression (TTP) nor with overall survival (OS) in 14 patients (p = 0.96 and 0.43, respectively). Likewise, McHugh et al. [30] investigated the effect of dexamethasone, a drug used to mitigate side effects of chemotherapy, on pemetrexed efficacy. They demonstrated that [¹⁸F]FLT could critically detect heterogeneity in dexamethasone sensitivity between tumours within individual patients. A comparison with [¹⁸F]FDG-PET was performed in 9 patients treated with neoadjuvant chemotherapy in a former study and categorised according to tumour size assessed by computed tomography [29]. Using [¹⁸F]FDG, anatomic responders showed significantly lower uptake than anatomic non-responders 2-3 weeks after treatment, while no significant difference was found between these subgroups using $[^{18}F]FLT$.

Radiotherapy

Preliminary data of studies that evaluated response to radiotherapy is more promising. Vera et al. [31] demonstrated significant changes in SUVmax in 5 patients treated with 46 Gy radiotherapy. This decline of $[^{18}F]$ FLT uptake was supported by another study (n = 20) which tested correlation with clinical results after treatment with carbon ion radiotherapy [32]. Baseline SUVmax

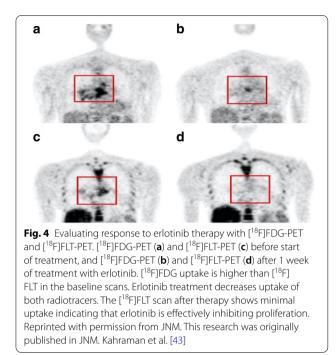
was predictive of response as patients who died or developed recurrence had significantly more [¹⁸F]FLT uptake than those did not (p = 0.007 and p = 0.008, respectively). Furthermore, primary tumours showed decreased SUVmean and SUVmax despite the absence of morphological change [33].

Chemoradiotherapy

Three studies have investigated the potential of [¹⁸F]FLT for assessing response to chemoradiotherapy. Response to chemoradiotherapy was firstly assessed in a small study (n = 5), which observed decreased $[^{18}F]FLT$ uptake in primary tumours after therapy [34]. A subsequent study (n = 20) found that $[^{18}F]FLT$ was more sensitive than [¹⁸F]FDG in assessing response to radical chemoradiation [35]. Median SUVmax were 14 and 6 at baseline and 10 and 3 two weeks post-therapy for [¹⁸F]FDG and [¹⁸F]FLT, respectively. The same group also studied correlation with clinical outcome in a larger cohort (n = 60) treated with 60 Gy radiotherapy combined with carboplatin and paclitaxel or cisplatin (32). Stable disease, as assessed using RECIST criteria, was associated with longer OS and progression-free survival (PFS) than patients with complete or partial response. This paradox could be ascribed to the weakening of tumouricidal effect of chemotherapy by suppressed proliferation [36].

Targeted therapy

Multiple studies have found high potential of [¹⁸F]FLT to predict LC response to targeted therapy. Yang et al. analysed the prediction of response to anti-angiogenic



agents in correlation with microvessel density (MVD) [37]. [¹⁸F]FLT was correlated significantly with MVD as reflected by CD105-MVD as well as clinical outcomes. Longer median survival times were observed in patients who had [18F]FLT false negative results (p = 0.012) and in patients with lower CD105-MVD (p = 0.046). Moreover, the ability of [¹⁸F]FLT to monitor pharmacodynamic effect therapy was examined by Scarpelli et al. with static (n = 14) and dynamic (n = 33) PET [38, 39]. In the static PET study, SUVmax was decreased -11% in cycle 1 treatment with the anti-angiogenic x-82. After administration of cycle 2 x-82 combined with the chemotherapy docetaxel on day 21, SUVmax was greatly decreased to -44% (34). Decline of cell tumour proliferation and vasculature were also exhibited 2 weeks after treatment with axitinib followed by an increase in washout period during the week of treatment break [39].

Assessment of epidermal growth factor receptor-tyrosine kinase inhibitors (EGFR-TKI) response with [¹⁸F]FLT is also an active area of research. Typical scans of [¹⁸F] FLT uptake before and after EGFR-TKI therapy in a LC patient responding to therapy are shown in Fig. 4. The value of [¹⁸F]FLT in this role is due to well-established clinical benefits of EGFR-TKI, as they are being used as first-line therapy in select NSCLC patients [47]. Three studies investigated the performance of [¹⁸F]FLT in assessing tumour progression in correlation with tumour size measured by computed tomography. Sohn et al. utilised [¹⁸F]FLT to predict response to gefitinib in 31 adenocarcinoma patients [40]. A significant difference (p < 0.001) was shown between responders and non-responders. Mileshkin et al. also evaluated [¹⁸F]FLT and [¹⁸F]FDG to monitor response in 51 NSCLC patients 2 and 8 weeks after treatment with erlotinib (first generation EGFR-TKI drug). 4 responders were assigned by computed tomography; all of them were classified responders by [¹⁸F]FDG and 3 of them by [¹⁸F]FLT [41]. In contrast, Zander et al. found that tumour size measurements correlated with changes in [¹⁸F]FDG uptake (p < 0.05) but not with [¹⁸F]FLT (p > 0.05) 6 weeks after erlotinib therapy [42]. Moreover, further clinical assessment of [¹⁸F]FLT role for early monitoring of response to newer generations of EGFR-TKI agents is still required in order to better understand their associated mechanisms of responsiveness and resistance [48].

The predictive role of clinical outcome using [¹⁸F]FLT uptake was also studied in EGFR-TKI studies and showed positive results. Sohn et al. demonstrated that [¹⁸F]FLT responders showed significantly longer TTP (p = 0.0041) than non-responders. Further, Scheffler et al. (n = 40) observed that [¹⁸F]FLT could prognostically stratify NSCLC patients treated with erlotinib, as patients with low uptake had significantly longer survival times (p = 0.027) than patients with high uptake [44]. Moreover, apart from one study with a small patient cohort (n = 15) [45], [¹⁸F]FLT uptake was correlated with PFS in multiple independent studies with larger cohorts [41–43]. However, in most studies OS did not correlate with [¹⁸F]FLT uptake [42, 45].

Discussion

This paper has reviewed the literature of clinical [¹⁸F]FLT applications in LC, and compared the uptake values of [¹⁸F]FLT and [¹⁸F]FDG in the various main LC sub-types. The review clearly shows that [¹⁸F]FLT is able to provide useful data to help diagnose, stage and monitor therapy response in LC. However, the value of using [¹⁸F]FLT-PET in some of these roles is stronger than in others. The paper also outlines some novel observations about the studies performed to date and provides some recommendations that could help to guide the design of prospective clinical studies with [¹⁸F]FLT in LC.

For diagnosis and staging, [¹⁸F]FLT showed better specificity and discriminative value between inflammatory and malignant LC than [¹⁸F]FDG. The superior specificity of [¹⁸F]FLT can be attributed to its highly selective uptake in highly proliferating (i.e. malignant) cells, providing better differentiation of malignant from benign tumours. [¹⁸F]FDG exhibits a slightly lower specificity than [¹⁸F]FLT due to the high metabolic activity of some benign lesions and inflammatory cells which use glucose as the main substrate for energy production [19]. On the other hand, [¹⁸F]FLT shows significantly lower uptake and, thus, sensitivity than $[^{18}F]FDG$ in LC. The lower sensitivity of [18F]FLT is in concordance with a previous systematic review and meta-analysis study which also deduced that [¹⁸F]FLT is a more specific but less sensitive radiotracer than [18F]FDG [49]. The low sensitivity of [18F]FLT appears to be the reason behind its understaging of tumours compared to [¹⁸F]FDG. This limitation makes it unable to replace [¹⁸F]FDG in these roles, although it may be able to complement $[^{18}F]FDG$ in some situations. The relatively low uptake of [¹⁸F]FLT in cancer cells is partly due to the fact that cells do not express a specialised transporter to facilitate its transport across the cell membrane; [¹⁸F]FLT either enters cells via the general nucleoside transporter hENT1 or via passive diffusion [50]. Furthermore, highly proliferating cells such as bone marrow act as a sink, reducing the amount of [¹⁸F]FLT available in the body to be taken up by tumours [51]. In contrast, cancer cells tend to take up relatively high levels of [¹⁸F]FDG. This is due to multiple factors. Firstly, they express specialised receptors such as GLUT1, which facilitate the active transport of glucose or [¹⁸F]FDG across the cell membrane, and partly due to the Warburg effect, which results in relatively high glucose metabolism in tumour cells. Secondly, whilst most cancer cells are highly metabolically active only a proportion of them are proliferating at any time. Finally, inflammatory cells are usually present in malignant tumours, which will further increase [¹⁸F]FDG uptake in regions of interest.

One interesting suggestion is that using [¹⁸F]FLT and ^{[18}F]FDG-PET in combination could potentially provide better diagnostic performance than the individual tracers. Indeed, this strategy showed higher sensitivity and specificity in diagnosing pulmonary lesions [21]. However, this benefit is offset by the higher costs associated with performing two PET scans and the increased radiation burden imposed by this dual tracer approach. There is an effective dose equivalent of 0.031 mSv/MBq and 0.029mSv/MBg for [¹⁸F]FLT and [¹⁸F]FDG, respectively [52]. When 2 doses of 400 MBg were administered by Tian et al. [21] this resulted in a total radiation dose per patient of approximately 24 mSv (12.4 from [¹⁸F] FLT + 11.6 from [¹⁸F]FDG) within a week. Owing to the increased radiation dose, the dual tracer strategy should be valued favourably only for patients with an equivocal diagnosis where the benefit of more confident diagnosis could brought to the patient, taking into account the patent's clinical characteristics.

An analysis of LC studies to date confirmed that there is significantly lower $[^{18}F]FLT$ uptake than $[^{18}F]FDG$ uptake in all histotypes tested (Fig. 3). This result was expected given the defined uptake mechanism for $[^{18}F]FDG$ in tumour cells combined with the Warburg effect, which results in tumour cells utilising high amounts of glucose.

A comparison of [¹⁸F]FDG-SUVmax detected no significant difference between the 4 sub-types of LC (p > 0.05). However, the statistical power of this analysis is limited by the number of SCLC studies performed to date and the relative high variation in uptake values between studies in the other tumour types. Publication of additional LC datasets is required to confirm if there is any clear differences in [¹⁸F]FDG uptake between LC sub-types. In contrast, a comparison of [¹⁸F]FLT-SUVmax between the different histotypes observed that [¹⁸F]FLT-SUVmax was significantly lower in SCLC than in SqCC (p < 0.05). This is a new observation which suggests that there may be something subtly different between these LC sub-types. There are 3 potential straightforward explanations for this observation. The first possibility is that SCLC proliferates at a slower rate. This seems unlikely given that SCLC is known to be an aggressive sub-type that grows quickly. The second possibility is that there is less cellular [¹⁸F]FLT uptake into SCLC, either because these tumours have limited access to [18F]FLT in the blood or because less radiotracer can cross the plasma-membrane, perhaps suggesting that the cells may express fewer hENT transporters. The third possibility is that SCLC could rely more than SqCC on the *de novo* pathway to synthesise nucleotides rather than using the salvage pathway [53], resulting in less [¹⁸F]FLT uptake into cells via hENT transporters. To our knowledge neither of the latter possibilities have been investigated to date.

Evaluating the potential of [¹⁸F]FLT-PET for assessing early response to therapy is a logical step, given the clear utility of [¹⁸F]FDG-PET in this role in LC. The rationale for using [¹⁸F]FLT-PET to assess therapies that selectively affect proliferation is especially compelling, as these treatments may inhibit proliferation without affecting either metabolic rate or causing tumour shrinkage [54]. Only 4 out of the 18 LC response assessment studies identified in this review did not observe some utility for $[^{18}F]$ FLT in this role [28, 29, 36, 42]. One of these studies [28] assessed a chemotherapeutic agent that is antagonist to thymidylate synthase. This may impact TK1, resulting in poor correlation between [¹⁸F]FLT uptake and clinical outcomes [53]. Whilst another study [29], which studied platinum-based chemotherapy, showed no correlation with CT response. The reason for this discrepancy is unknown. These findings suggest that the complex effect of cytotoxic therapeutics on cell signalling pathways needs to be fully considered to understand how each agent may affect [¹⁸F]FLT uptake into cells. Indeed, a previous systematic review that evaluated the role of [¹⁸F]FLT as a measure of therapy response in different tumours also concluded that [18F]FLT-PET is not as useful as [¹⁸F]FDG-PET for assessing chemotherapeutic response in NSCLC [55]. However, studies evaluating these types of therapies are still scarce and findings from more studies with larger patient cohorts might be more conclusive. A chemoradiotherapy study which employed ^{[18}F]FLT-PET to evaluate therapy response observed that stable disease measurements compared with complete or partial response based on CT was associated with longer OS and progression-free survival (PFS) [36]. This paradox could be ascribed to the weakening of tumouricidal effect of chemotherapy by suppressed proliferation [36]. Finally, a study evaluating [¹⁸F]FLT-PET to assess EGFR-TKI response found no correlation with CT measurements [42]. However, this observation may be due to utilisation of size measurements to assess EGFR-TKI; change in tumour size may be delayed or may not occur at all with this type of therapy, creating a discrepancy between these two different metrics [54].

The most promising application for [¹⁸F]FLT is assessing response to targeted agents that selectively inhibit cell proliferation. In this role [¹⁸F]FLT is utilised as a precise tool for detecting the effects of antiproliferative therapies. Apart from one study with a small patient cohort (n = 15) [45], $[^{18}F]FLT$ uptake correlated with PFS in multiple independent studies with larger cohorts [41-43]. Interestingly, in most studies OS did not correlate with [¹⁸F]FLT uptake [42, 45]. This may be due to the subtly different metrics captured by PFS and OS. The OS statistics may be somewhat compromised by the limited number of patients recruited in most of these studies and/or the limited follow up time. Indeed, the study with the largest patient group (n = 51) and follow up time observed correlation between [18F]FLT uptake and OS 8 weeks after treatment; the longest follow up time in other studies was only 6 weeks [41, 42]. Although OS is a gold standard measure to demonstrate clinical efficacy, larger patient numbers and longer follow up are required to establish [18F]FLT as a reliable measure of this endpoint [56]. PFS is more advantageous than OS in that it assesses both stable disease and responsive disease. This renders it a more reasonable clinical measure for targeted therapies, which often benefit patients mainly through prolonged stable disease rather than tumour shrinkage [57]. The assessment of pharmacodynamic endpoints is another extremely important function for [¹⁸F]FLT. Scarpelli et al. [38, 39] demonstrated that the increase of $[^{18}F]$ FLT uptake is apparently caused by the washout period during treatment. This type of study, which evaluates the pharmacodynamic endpoints, is of great importance to develop better understanding of drug resistance mechanisms, as it helps clinicians to tailor more effective treatments and to minimise systematic toxicity caused by ineffective drugs [38]. Together, these preliminary findings suggest that [18F]FLT is a useful tool for evaluating response to targeted therapies with anticipated cytostatic effects, but further studies with different targeted agents, larger patient cohorts and longer follow ups are warranted to fully understand its potential in this role.

The studies mentioned above have multiple limitations. One limitation is that most studies are singlecentre observational studies with small patient cohorts. This makes it difficult to know how readily the study findings would translate to other research centres or if similar results would be obtained using larger patient cohorts. While most studies used a relatively similar imaging protocol, there were no standard instructions for patient preparation. Some studies encouraged patients to drink water before imaging, which would help to reduce background radiation, whereas others did not. In addition, there were differences between studies in tracer dose administered, which is likely to affect the signal to background ratio. Most studies utilised static scans, with similar times between tracer injection and imaging. However, the use of different reconstruction techniques and metrics for assessing uptake (e.g. FLTmax vs FLTmean) would result in subtly different values being calculated from the same dataset. It is important that methods are optimised and standardised, so that the results obtained in one study can be directly compared with those from another, and so that multi-centre studies can generate equivalent data at all participating centres. This issue has been recognised within the PET community and has resulted in core labs being set up to ensure quality control for PET clinical trials [58]. The limited number of studies evaluating LC response to various types of treatment affects the conclusiveness of study findings. Moreover, size-based measurements were used as a standard reference in studies evaluating responses to targeted therapy although tumour size would not be affected with this type of therapy. OS was also used as the gold standard for assessing clinical outcome in studies evaluating responses to targeted therapy although it showed limitations compared with surrogate metrics such as PFS. Therefore, extra attention is also needed to ensure that the most suitable metrics of clinical outcome are used to study such therapies with [¹⁸F]FLT-PET.

Conclusions

1. Overall, [¹⁸F]FLT seems to have better specificity in diagnosis and staging but lower sensitivity with significantly lower uptake than [¹⁸F]FDG. This significant difference was also observed after comparing uptake values of both tracers in the main histotypes of lung cancer that showed consistence with established histological patterns of cell proliferation in

both tracers. Therefore, $[^{18}F]FLT$ cannot be considered superior to $[^{18}F]FDG$ for diagnosis and staging.

- 2. [¹⁸F]FLT uptake values of SCLC is considerably lower than SqCC. This indicates that SCLC may not be suitable for [¹⁸F]FLT imaging studies, due to limited radiotracer uptake.
- 3. [¹⁸F]FLT can provide good predictive values in patients undergoing radiotherapy or chemoradio-therapy and looks especially useful for assessing early response to targeted therapies.
- 4. [¹⁸F]FLT uptake values correlate well with TTP and PFS, but less well with size measurements based on computed tomography and OS.
- 5. [¹⁸F]FLT-PET tends to be promising in preliminary results of pharmacodynamic endpoints of targeted therapy predicting potential resistance which could permit better individualisation of treatment plans.

Recommendations

- 1. More suitable reference standards such as histopathology for studies assessing response to targeted therapies are recommended for future studies.
- 2. TTP or PFS appear to be suitable clinical endpoints for [¹⁸F]FLT response assessment studies with targeted agents.
- 3. Studies should carefully consider which LC sub-type is recruited for imaging studies with [¹⁸F]FLT, as SCLC sub-type appears to have limited uptake.
- 4. More [¹⁸F]FLT studies are needed, to further evaluate the potential for this radiotracer to assess early response to therapy. This should include studies which evaluate different targeted therapies, studies which use standardised imaging protocols, multicentre studies, and studies with larger patient cohorts and longer follow up times.

Abbreviations

[¹¹C]thymidine: ¹¹C-labelled thymidine; [¹⁸F]FBAU: 1-(2-deoxy-2-[¹⁸F]fluoro-βd-arabinofuranosyl)-5-bromouracil; [¹⁸F]FDG: [¹⁸F]fluorodeoxyglucose; [¹⁸F]FLT: 3'-deoxy-3'-[¹⁸F]fluorothymidine; [¹⁸F]FMAU: 2'-deoxy-2'-[¹⁸F]fluoro-5-methyl-1-β-d-arabinofuranosyluracil; ADC: Adenocarcinoma; EGFR-TKI: Epidermal growth factor receptor- tyrosine kinase inhibitor; LC: Lung cancer; LCC: Large cell lung carcinoma; MVD: Microvessel density; n: Number of patients; NSCLC: Non-small cell lung cancer; OS: Overall survival; PET: Positron emission tomography; PFS: Progression free survival; SCLC: Small cell lung cancer; SPF: S-phase fraction; SqCC: Squamous cell carcinoma; SUV: Standardised uptake value; SUVmax: Maximum standardised uptake value; SUVmean: Mean standardised uptake value; SUVpeak: Standardised uptake value peak; TK1: Thymidine kinase 1; TK2: Thymidine kinase 2; TNM: Tumour, nodal involvement and metastases; TTP: Time to progression.

Authors' contributions

B.A. implemented the literature search, data Curation and formal analysis, and wrote the first draft of the manuscript and edited various versions of manuscript; B.A. and I.N.F. implemented data curation and formal analysis. S.D. and I.N.F. were major contributors in revising the manuscript. All authors read and approved the final manuscript.

Funding

This review was supported by funding provided by the Royal Embassy of Saudi Arabia, Cultural Bureau in London, UK. We also gratefully acknowledge Friends of Anchor for their generous financial support of our research.

Availability of data and materials

The datasets used and/or analyses during the current study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate Not applicable

tot applicable

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

Author details

¹Diagnostic Radiology Department, College of Applied Medical Sciences, Jazan University, Al Maarefah Rd, POB 114, Jazan 45142, Saudi Arabia. ²Institute of Medical Sciences, University of Aberdeen, Aberdeen AB25 2ZD, UK.

Received: 28 January 2021 Accepted: 2 June 2021 Published online: 02 July 2021

References

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA A Cancer J Clin 68(6):394–424
- 2. Kadara H, Scheet P, Wistuba LI, Spira AE (2016) Early events in the molecular pathogenesis of lung cancer. Am Assoc Cancer Res 9(7):518–527
- 3. Rinaldi S, Berardi R (2017) Lung cancer prognosis: Can histological patterns and morphological features have a role in the management of lung cancer patients? Ann Transl Med 5(17)
- Ruben JD, Ball DL (2012) The efficacy of PET staging for small-cell lung cancer: a systematic review and cost analysis in the Australian setting. J Thorac Oncol 7(6):1015–1020
- Chalkidou A, Landau DB, Odell EW, Cornelius VR, O'Doherty MJ, Marsden PK (2012) Correlation between Ki-67 immunohistochemistry and 18F-Fluorothymidine uptake in patients with cancer: a systematic review and meta-analysis. Eur J Cancer 48(18):3499–3513
- Shen G, Ma H, Pang F, Ren P, Kuang A (2018) Correlations of 18F-FDG and 18F-FLT uptake on PET with Ki-67 expression in patients with lung cancer: a meta-analysis. Acta Radiol 59(2):188–195
- Tabata K, Tanaka T, Hayashi T et al (2014) Ki-67 is a strong prognostic marker of non-small cell lung cancer when tissue heterogeneity is considered. BMC Clin Pathol 14(1):1–8
- Al-Jahdali H, Khan AN, Loutfi S, Al-Harbi AS (2012) Guidelines for the role of FDG-PET/CT in lung cancer management. J Infect Public Health 5(5 Suppl 1):35–40
- Deppen SA, Blume JD, Kensinger CD et al (2014) Accuracy of FDG-PET to diagnose lung cancer in areas with infectious lung disease: a meta-analysis. JAMA 312(12):1227–1236
- Buck AK, Herrmann K, Shen C, Dechow T, Schwaiger M, Wester H (2009) Molecular imaging of proliferation in vivo: positron emission tomography with [18F]fluorothymidine. Methods 48(2):205–215

- Alegre MM, Robison RA, O'Neill KL (2013) Thymidine kinase 1: a universal marker for cancer. Cancer Clin Oncol 2(1):159–167
- 12. Alegre MM, Robison RA, O'Neill KL (2012) Thymidine kinase 1 upregulation is an early event in breast tumor formation. J Oncol 2012:575647. https://doi.org/10.1155/2012/575647
- Sun H, Sloan A, Mangner TJ, Vaishampayan U, Muzik O, Collins JM et al (2005) Imaging DNA synthesis with [18F]FMAU and positron emission tomography in patients with cancer. Eur J Nucl Med Mol Imaging 32(1):15–22
- 14. Vesselle H, Grierson J, Muzi M, Pugsley JM, Schmidt RA, Rabinowitz P et al (2002) In vivo validation of 3'deoxy-3'-[18F]fluorothymidine ([18F]FLT) as a proliferation imaging tracer in humans: correlation of [18F]FLT uptake by positron emission tomography with Ki-67 immunohistochemistry and flow cytometry in human lung tumors. Clin Cancer Res 8(11):3315–3323
- Cobben DCP, Elsinga PH, Hoekstra HJ, Suurmeijer AJH, Vaalburg W, Maas B et al (2004) Is 18F–3'-fluoro-3'-deoxy-L-thymidine useful for the staging and restaging of non-small cell lung cancer? J Nucl Med 45(10):1677–1682
- Buck AK, Schirrmeister H, Hetzel M, Von Der Heide M, Halter G, Glatting G et al (2002) 3-Deoxy-3-[18F]fluorothymidine-positron emission tomography for noninvasive assessment of proliferation in pulmonary nodules. Cancer Res 62(12):3331–3334
- Buck AK, Hetzel M, Schirrmeister H, Halter G, Möller P, Kratochwil C et al (2005) Clinical relevance of imaging proliferative activity in lung nodules. Eur J Nucl Med Mol Imaging 32(5):525–533
- Wang F, Tan Y, Gu X, Li T, Lu G, Liu G et al (2016) Comparison of positron emission tomography using 2-[18F]-fluoro-2-deoxy-D-glucose and 3-deoxy-3-[18F]-fluorothymidine in lung cancer imaging. Chin Med J 129(24):2926–2935
- Yamamoto Y, Nishiyama Y, Kimura N, Ishikawa S, Okuda M, Bandoh S et al (2008) Comparison of 18F-FLT PET and 18F-FDG PET for preoperative staging in non-small cell lung cancer. Eur J Nucl Med Mol Imaging 35(2):236–245
- Yang W, Zhang Y, Fu Z, Yu J, Sun X, Mu D et al (2010) Imaging of proliferation with 18F-FLT PET/CT versus 18F-FDG PET/CT in non-small-cell lung cancer. Eur J Nucl Med Mol Imaging 37(7):1291–1299
- Tian J, Yang X, Yu L, Chen P, Xin J, Ma L et al (2008) A multicenter clinical trial on the diagnostic value of dual-tracer PET/CT in pulmonary lesions using 3'-deoxy-3'-18F- fluorothymidine and 18F-FDG. J Nucl Med 49(2):186–194
- Buck AK, Halter G, Schirrmeister H, Kotzerke J, Wurziger I, Glatting G et al (2003) Imaging proliferation in lung tumors with PET: 18F-FLT versus 18F-FDG. J Nucl Med 44(9):1426–1431
- Dittmann H, Dohmen BM, Paulsen F, Eichhorn K, Eschmann SM, Horger M et al (2003) [18F]FLT PET for diagnosis and staging of thoracic tumours. Eur J Nucl Med Mol Imaging 30(10):1407–1412
- Yap CS, Czernin J, Fishbein MC, Cameron RB, Schiepers C, Phelps ME et al (2006) Evaluation of thoracic tumors with 18F-fluorothymidine and 18F-fluorodeoxyglucose-positron emission tomography. Chest 129(2):393–401
- Yamamoto Y, Nishiyama Y, Ishikawa S, Nakano J, Chang SS, Bandoh S et al (2007) Correlation of 18F-FLT and 18F-FDG uptake on PET with Ki-67 immunohistochemistry in non-small cell lung cancer. Eur J Nucl Med Mol Imaging 34(10):1610–1616
- Xu X, Nie L, Yao Y, Tu Y, Zhou J (2016) Pre-radiotherapy assessment of nonsmall cell lung cancer with 18F-FLT PET/CT or 18F-FDG PET/CT. Int J Clin Exp Med 9(6):9252–9260
- 27. Vesselle H, Grierson J, Muzi M, Pugsley JM, Schmidt RA, Rabinowitz P et al (2002) In vivo validation of 3'deoxy-3'-[18F]fluorothymidine ([18F]FLT) as a proliferation imaging tracer in humans: correlation of [18F]FLT uptake by positron emission tomography with Ki-67 immunohistochemistry and flow cytometry in human lung tumors. Clin Cancer Res 8(11):3315–3323
- Frings V, van der Veldt AAM, Boellaard R, Herder GJM, Giovannetti E, Honeywell R et al (2013) Pemetrexed induced thymidylate synthase inhibition in non-small cell lung cancer patients: a pilot study with 3'-deoxy-3'-[18f] fluorothymidine positron emission tomography. PLoS One 8(5)
- Crandall JP, Tahari AK, Juergens RA, Brahmer JR, Rudin CM, Esposito G et al (2017) A comparison of FLT to FDG PET/CT in the early assessment of chemotherapy response in stages IB–IIIA resectable NSCLC. EJNMMI Res 7(1).

- McHugh CI, Thipparthi MR, Lawhorn-Crews JM, Polin L, Gadgeel S, Akoury J et al (2018) Using radiolabeled 3'-Deoxy-3'-18F-Fluorothymidine with PET to monitor the effect of dexamethasone on non-small cell lung cancer. J Nucl Med 59(10):1544–1550
- 31. Vera P, Bohn P, Edet-Sanson A, Salles A, Hapdey S, Gardin I et al (2011) Simultaneous positron emission tomography (PET) assessment of metabolism with 18F-fluoro-2-deoxy-d-glucose (FDG), proliferation with 18F-fluoro-thymidine (FLT), and hypoxia with 18fluoro- misonidazole (F-miso) before and during radiotherapy in patients with non-small-cell lung cancer (NSCLC): a pilot study. Radiother Oncol 98(1):109–116
- Saga T, Koizumi M, Inubushi M, Yoshikawa K, Tanimoto K, Fukumura T et al (2011) PET/CT with 3'-deoxy-3'-[18F]fluorothymidine for lung cancer patients receiving carbon-ion radiotherapy. Nucl Med Commun 32(5):348–355
- Trigonis I, Koh PK, Taylor B, Tamal M, Ryder D, Earl M et al (2014) Early reduction in tumour [18F]fluorothymidine (FLT) uptake in patients with non-small cell lung cancer (NSCLC) treated with radiotherapy alone. Eur J Nucl Med Mol Imaging 41(4):682–693
- 34. Everitt S, Hicks RJ, Ball D, Kron T, Schneider-Kolsky M, Walter T et al (2009) Imaging cellular proliferation during chemo-radiotherapy: a pilot study of serial 18F-FLT positron emission tomography/computed tomography imaging for non-small-cell lung cancer. Int J Radiat Oncol Biol Phys 75(4):1098–1104
- Everitt SJ, Ball DL, Hicks RJ, Callahan J, Plumridge N, Collins M et al (2014) Differential 18F-FDG and 18F-FLT uptake on serial PET/CT imaging before and during definitive chemoradiation for non-small cell lung cancer. J Nucl Med 55(7):1069–1074
- 36. Everitt S, Ball D, Hicks RJ, Callahan J, Plumridge N, Trinh J et al (2017) Prospective study of serial imaging comparing fluorodeoxyglucose positron emission tomography (PET) and fluorothymidine PET during radical chemoradiation for non-small cell lung cancer: reduction of detectable proliferation associated with worse survival. Int J Radiat Oncol Biol Phys 99(4):947–955
- Yang W, Zhang Y, Fu Z, Sun X, Mu D, Yu J (2012) Imaging proliferation of 18F-FLT PET/CT correlated with the expression of microvessel density of tumour tissue in non-small-cell lung cancer. Eur J Nucl Med Mol Imaging 39(8):1289–1296
- Scarpelli M, Rampurwala M, Eickhoff J, Carmichael L, Heideman J, Binger K et al (2018) Pharmacodynamic study using FLT PET/CT in advanced solid malignancies treated with a sequential combination of X-82 and docetaxel. Cancer Chemother Pharmacol 82(2):211–219
- Scarpelli M, Simoncic U, Perlman S, Liu G, Jeraj R (2018) P Dynamic 18F-FLT PET imaging of spatiotemporal changes in tumor cell proliferation and vasculature reveals the mechanistic actions of anti-angiogenic therapy. Phys Med Biol 63(15).
- 40. Sohn H, Yang Y, Ryu J, Seung JO, Ki Cl, Dae HM et al (2008) [18 F]fluorothymidine positron emission tomography before and 7 days after gefitinib treatment predicts response in patients with advanced adenocarcinoma of the lung. Clin Cancer Res 14(22):7423–7429
- 41. Mileshkin L, Hicks RJ, Hughes BGM, Mitchell PLR, Charu V, Gitlitz BJ et al (2011) Changes in18F-fluorodeoxyglucose and 18F-fluorodeoxythymidine positron emission tomography imaging in patients with non-small cell lung cancer treated with erlotinib. Clin Cancer Res 17(10):3304–3315
- Zander T, Scheffler M, Nogova L, Kobe C, Engel-Riedel W, Hellmich M et al (2011) Early prediction of nonprogression in advanced non-small-cell lung cancer treated with erlotinib by using [18F] fluorodeoxyglucose and [18F] fluorothymidine positron emission tomography. J Clin Oncol 29(13):1701–1708
- 43. Kahraman D, Scheffler M, Zander T, Nogova L, Lammertsma AA, Boellaard R et al (2011) Quantitative analysis of response to treatment with erlotinib in advanced non-small cell lung cancer using 18F-FDG and 3'-deoxy-3'-18f-fluorothymidine PET. J Nucl Med 52(12):1871–1877
- 44. Scheffler M, Zander T, Nogova L, Kobe C, Kahraman D, Dietlein M et al (2013) Prognostic impact of [18F]fluorothymidine and [18F]fluoro-dglucose baseline uptakes in patients with lung cancer treated first-line with erlotinib. PLoS One 8(1)
- 45. Bhoil A, Singh B, Singh N, Kashyap R, Watts A, Sarika S et al (2014) Can 3'-deoxy-3'-18F-fluorothymidine or 2'-deoxy-2'-18F-fluoro-d-glucose PET/CT better assess response after 3-weeks treatment by epidermal growth factor receptor kinase inhibitor, in non-small lung cancer patients? Preliminary results Hell J Nucl Med 17(2):90–96

- 46. Yap TA, Vidal L, Adam J, Stephens P, Spicer J, Shaw H et al (2010) Phase I trial of the irreversible EGFR and HER2 kinase inhibitor BIBW 2992 in patients with advanced solid tumors. J Clin Oncol 28(25):3965–3972
- 47. Chen G, Kronenberger P, Teugels E, Umelo IA, De Grève J (2012) Targeting the epidermal growth factor receptor in non-small cell lung cancer cells: The effect of combining RNA interference with tyrosine kinase inhibitors or cetuximab. BMC Med 10
- Zannetti A, Iommelli F, Speranza A, Salvatore M, Del Vecchio S (2012) 3'-Deoxy-3'- 18F-fluorothymidine PET/CT to guide therapy with epidermal growth factor receptor antagonists and Bcl-x L inhibitors in non-small cell lung cancer. J Nucl Med 53(3):443–450
- Wang Z, Wang Y, Sui X, Zhang W, Shi R, Zhang Y et al (2015) Performance of FLT-PET for pulmonary lesion diagnosis compared with traditional FDG-PET: A meta-analysis. Eur J Radiol 84(7):1371–1377
- Plotnik DA, Emerick LE, Krohn KA, Unadkat JD, Schwartz JL (2010) Different modes of transport for 3H-thymidine, 3H-FLT, and 3H-FMAU in proliferating and nonproliferating human tumor cells. J Nucl Med 51(9):1464–1471
- Agool A, Schot BW, Jager PL, Vellenga E (2006) 18F-FLT PET in hematologic disorders: a novel technique to analyze the bone marrow compartment. J Nucl Med 47(10):1592–1598
- Vesselle H, Grierson J, Peterson LM, Muzi M, Mankoff DA, Krohn KA (2003) 18F-fluorothymidine radiation dosimetry in human PET imaging studies. J Nucl Med 44(9):1482–1488

- Schelhaas S, Heinzmann K, Bollineni VR, Kramer GM, Liu Y, Waterton JC et al (2017) Preclinical applications of 3'-deoxy-3'-[18F]fluorothymidine in oncology—a systematic review. Theranostics 7(1):40–50
- Fernandes RS, de Aguiar FC, Soares DCF, Maffione AM, Townsend DM, Rubello D et al (2017) The role of radionuclide probes for monitoring antitumor drugs efficacy: a brief review. Biomed Pharmacother 95:469–476
- Bollineni VR, Kramer GM, Jansma EP, Liu Y, Oyen WJG (2016) A systematic review on [18F]FLT-PET uptake as a measure of treatment response in cancer patients. Eur J Cancer 55:81–97
- Gill S, Berry S, Biagi J, Butts C, Buyse M, Chen E et al (2011) Progressionfree survival as a primary endpoint in clinical trials of metastatic colorectal cancer. Curr Oncol 18(SUPPL. 2):S5–S10
- Villaruz LC, Socinski MA (2013) The clinical viewpoint: Definitions, limitations of RECIST, practical considerations of measurement. Clin Cancer Res 19(10):2629–2636
- NCRI PET Core Lab—Structure. http://www.ncri-pet.org.uk/structure.php. Accessed 15 April 2021

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- ► Rigorous peer review
- Open access: articles freely available online
- ► High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at > springeropen.com